



Antimicrobial Potential of *Coleus Atropurpureus* and its Metabolites Content: A Literature Review

Ayu Suci Pratiwi^{1*}, Debby Handayati Harahap², Erizka Rivani³

¹Program Pascasarjana Ilmu Biomedik, Fakultas Kedokteran, Universitas Sriwijaya, Indonesia

²Departemen Farmakologi, Fakultas Kedokteran, Universitas Sriwijaya, Indonesia

³Departemen Mikrobiologi, Fakultas Kedokteran, Universitas Sriwijaya, Indonesia

*Penulis Korespondensi: dr.ayusucipratiwi@gmail.com

Article History:

Received: 20 August 2025;

Revised: 04 September 2025;

Accepted: 18 September 2025;

Published: 04 October 2025.

Keywords: Antimicrobial Activity; *Coleus Atropurpureus*; Drug-Resistant Bacteria; Phytochemicals; Synergistic Effects.

Abstract: The increasing prevalence of multidrug-resistant bacterial infections underscores the urgent need for new antimicrobial agents, especially those derived from plant-based sources. *Coleus atropurpureus*, a medicinal plant used extensively in Asia and Oceania, contains various bioactive compounds, including flavonoids, terpenoids, and phenolic acids, which have demonstrated antimicrobial properties. This review examines studies published between 2015 and 2025, highlighting the antibacterial activity of ethanolic and methanolic extracts of *C. atropurpureus*, particularly against methicillin-resistant *Staphylococcus aureus* (MRSA). Phytochemical analysis reveals that these extracts exert their antimicrobial effects through multiple mechanisms, including the disruption of microbial membranes, enzyme inhibition, the induction of reactive oxygen species, and immunomodulatory actions. Furthermore, the extracts have shown synergistic effects when combined with antibiotics, suggesting their potential as adjunct therapies. The evidence gathered supports the therapeutic potential of *C. atropurpureus* as a natural antimicrobial agent, offering a promising alternative to conventional antibiotics in combating drug-resistant infections.

1. INTRODUCTION

Opportunistic bacterial infection caused by multidrug-resistant pathogens have become a growing public health concern worldwide. The advent of antimicrobial resistance has added significant impact of infectious diseases, in number of infectins, as well as added healthcare costs. These concerns prompted the World Health Organization (WHO) to launch a Global Action Plan on antimicrobial resistance in 2015 (Organization 2015).

The rising incidence of drug-resistant pathogens highlights an urgent need to discover and isolate new medicinal compounds from plants. According to WHO, an estimated 80% of the developing countries still rely on the use of traditional medicines derived from plants (Organization 1978). WHO has also recommended the names of over 20.000 medicinal plant species as one of the potential sources for developing new drugs. The use of and search for drugs derived from plants have accelerated in recent years. Medicinal plant components could

provide novel approaches as one of the potential sources of new antimicrobe. While 25 to 50% of current pharmaceuticals are derived from plants, none are used as antibiotics (Vaou et al. 2021).

Coleus atropurpureus is a member of the Lamiaceae family, known as a decorative plant and traditional medicinal applications found in Southeast Asia, East Asia, and the Australasia and Oceania regions.(Büttner 2001) It is used for treating conditions such as skin infections, ear and eye infections, digestive disturbances, to inflammation.(Quisumbing 1978; Roosita et al. 2008; Quattrocchi 2012; Plants of the World Online 2023).

The genus *Coleus* is known with notable biological properties, such as antioxidant, antifungal, antiparasitic, anti-inflammatory, anti-diabetic, anti-cancer, and antibacterial effects (Ridwan and Ayunita 2007; Zakaria et al. 2008; Namsa et al. 2009; Arumugam et al. 2016; Barbosa et al. 2023). Specifically, parts and content of *C. atropurpureus* is abundant in a wide variety of metabolites content, such as flavonoids, phenolic acids, alkaloids, saponins, tannins, and terpenes (Lamprecht et al. 1975; Devriese et al. 1988; Mu et al. 1996; Ragasa et al. 2001; Lukhoba et al. 2006; Ito et al. 2018) which have been reported to have antimicrobial properties.

To date, no narrative review has comprehensively summarized the bioactive mechanisms and potential clinical application of *Coleus atropurpureus* as an antimicrobial agent. This review aims to summarize the antimicrobial efficacy of *Coleus atropurpureus* and highlights the role of its metabolite content based on existing in vivo studies. The most relevant studies regarding the antimicrobial activity of *C. atropurpureus* plant extract, underlying mechanisms of action, and the challenges and future perspectives of the medicinal plant derived antibiotics are analyzed in this review.

2. METHOD

A comprehensive literature search was carried out across the databases PubMed, Scopus, Science Direct, and Google Scholar search engine. The terms used in the searches included, “phytochemicals”, and “antimicrobial” of “*Coleus*, *Plectranthus*, *Solenostemon atropurpureus* or *scutellarioides* or *blumei*”, among others related to the subject of the review. “AND” or “OR” Boolean operators were used depending on the combination of terms. In vivo studies, in vitro studies, and research articles were included. Inclusion criteria for this review were (1) studies published within the last 10 years (2015-2025), (2) articles in English or Bahasa Indonesia, and (3) accessible full text. Only studies that gave details on the mechanism of action, reliable data on antimicrobial activity, and the origin of the agents were selected for this review.

3. RESULT AND DISCUSSION

Result

Coleus atropurpureus

Coleus atropurpureus (or *Coleus scutellarioides*, *Solenostemon scutellarioides*, *Plectranthus scutellarioides*, *Coleus blumei*, *Plectranthus blumei*)(Paton et al. 2004) is a decorative plant with medicinal benefits belonging to the Lamiaceae family, Equisetopsida class, of the Streptophyta phylum. *Plectranthus scutellarioides* is distributed across tropical and subtropical regions, including the Bismarck Archipelago, New Guinea, the Solomon Islands, and Vanuatu in Oceania; the Philippines, Indonesia (Java, Sumatra, Sulawesi, Maluku, and the Lesser Sunda Islands), Malaysia, Thailand, Vietnam, Cambodia, Laos, Myanmar, and Singapore in Southeast Asia; southern China and Taiwan in East Asia; as well as parts of northern and western Australia, including the Northern Territory, Queensland, and Western Australia.(Plants of the World Online 2023) It is a subshrub and grows primarily in the wet tropical biome. The leaves of this plant can be up to 7 to 11 cm in length and 4 to 6 cm in width, heart shaped.(Wiart 2006) The yellow color of *C. blumei* leaves is mainly due to flavonoids like luteolin and quercetin,(Moektiwardoyo et al. 2011) while anthocyanins are responsible for the red or purple hues;(Nguyen and Cin 2009) other key compounds include organic acids,(Bauer et al. 2002) abietane-type diterpenoids,(Mu et al. 1996; Ito et al. 2018; Kubinová et al. 2019) and triterpenes such as β -amyrin and daucosterol.(Thomas 2006) In vitro studies have shown that these compounds can be extracted from both the aerial parts (e.g. stem, leaves) and roots of the plant.(Levita et al. 2016; Ito et al. 2018; Astuti et al. 2019; Bismelah et al. 2022; Kowalczyk et al. 2024)

Antibacterial Activity

The majority of reviewed studies in Table 1 used methanolic or ethanolic leaf extracts, which consistently showed activity against both Gram-positive and Gram-negative bacteria. Its antimicrobial potential was found to be active against *S. aureus*,(Bismelah et al. 2019; Bismelah et al. 2022; Bismelah et al. 2025; Hanum et al. [no date]) *E. coli*, *P. aeruginosa*,(Kaunang et al. 2016) *B. subtilis*, and *C. albicans*,(Karo et al. 2018) achieving moderate antimicrobial activity of 20–40% effectiveness compared to standard antibiotics (chloramphenicol, tetracycline, and chlortrimazole).(Ragasa et al. 2001) The antimicrobial efficacy was generally stronger against Gram-positive bacteria (e.g. *S. aureus* than *E. coli*), possibly due to differences in cell wall permeability.

Table 1. Phytochemicals Isolated from *C. Atropurpureus* with Antimicrobial Activity.

Phytochemical	Tested microorganism	Extract	Activity	Author (Year)
Mixed metabolites (flavonoids, tannins, terpenoids, saponins)	<i>S. aureus</i>	Ethanol extract	ZOI: 13-14.56 mm (100 mg/mL) MIC: 1.56 mg/mL MBC: 3.1 mg/mL	Bismelah et al. (2019)
Flavonoid	<i>A. actinomycetemcomitans</i> , <i>P. gingivalis</i> , <i>P. intermedia</i> , <i>S. mitis</i> , <i>S. oralis</i> , <i>S. salivarius</i> , <i>S. sanguinis</i> , <i>T. forsythia</i> , <i>T. denticola</i>	Ethanol extract (quercetin-3-glucoside, quercitrin, quercetin 3-(6"-acetylglucoside), quercetin 3-O-acetylramnoside)	ZOI: 13-19 mm (100 mg/mL) ZOI: 10-22 mm (200 mg/mL) MIC: 1.56 mg/mL (aerobes) MIC: 3.12 mg/mL (anaerobes) MBC: 3.13 mg/mL (aerobes) MBC: 6.25 mg/mL (anaerobes)	Bismelah et al. (2022)
Mixed metabolites (flavonoids, tannins, terpenoids, saponins)	<i>A. viscosus</i>	Ethanol extract	ZOI: 17 mm (100 mg/mL) ZOI: 21 mm (200 mg/mL) MIC: 3.6 mg/mL MBC: 12.5 mg/mL	Bismelah et al. (2025)
N/A	<i>S. aureus</i> , <i>E. coli</i>	Ethanol extract	ZOI: 7.32-9.83 mm (400 mg/mL)	Hanum et al. (2022)
Abietane diterpene	<i>S. aureus</i> CCM 4750 (MRSA)	Methanol extract (Sincoetsin C, Scutellarioidone A, Spirocoleon 7-O-b-D-glucoside, 3-Hydroxyspirocoleon 7-O-b-D-glucoside)	MIC: 128 µg/mL (Sincoetsin C) MIC: 512 µg/mL (others)	Jurkaninová et al. (2019)
Mixed metabolites (alkaloid, polyphenol, flavonoid, monoterpenoid and sesquiterpenoid, steroid and triterpenoid, quinone)	<i>S. aureus</i> ATTC 25923, MRSA	Ethanol extract	ZOI: 10.3-11.5 mm (62.5 mg/mL) ZOI: 11.5-13.6 mm (125 mg/mL) ZOI: 13.3-15.1 mm (250 mg/mL) ZOI: 16.2-21.1 mm (500 mg/mL)	Mustarichie et al. (2022)

ZOI, zone of inhibition; MIC, minimum inhibitory concentration; MBC, minimum bactericidal concentration; MRSA, methicilin-resistant *Staphylococcus aureus*; N/A, not available.

Several studies have systematically evaluated the antimicrobial potential of *Coleus atropurpureus*, using different extracts, phytochemicals, and microbial targets. In vitro studies consistently shows that extracts from *C. atropurpureus*, particularly those prepared with methanol or ethanol, possess significant antimicrobial activity against a range of pathogens. Based on the results of the in vitro antibacterial culture tests, *C. atropurpureus* extract concentration of 10-40% can be categorized in moderate inhibition ability. The higher extract concentration, the more bacterial destruction is expected. The relatively low MIC/MBC values suggest that *C. atropurpureus* metabolites are not only bacteriostatic but also shows bactericidal activity. The extract likely interfered with the bacterial cell division process, leading to distorted and irregular damage to the peptidoglycan layer and the cell membrane (Bismelah et al. 2025). Across all studies, *S. aureus* and MRSA were consistently more susceptible than Gram-negative bacteria such as *E. coli*, which reflects the typical resistance of Gram-negative strains due to the absence of an outer lipopolysaccharide protective outer membrane.

The effectiveness of a plant extract's antimicrobial properties is influenced by the solvent used during the extraction process. Specifically, organic solvents tend to be more effective at extracting the phytochemicals responsible for antimicrobial activity compared to water-based (aqueous) extracts (DAR 2016; Bismelah et al. 2025). It has been established that ethanol and methanol extracts contain more secondary metabolites and less impurities than water.

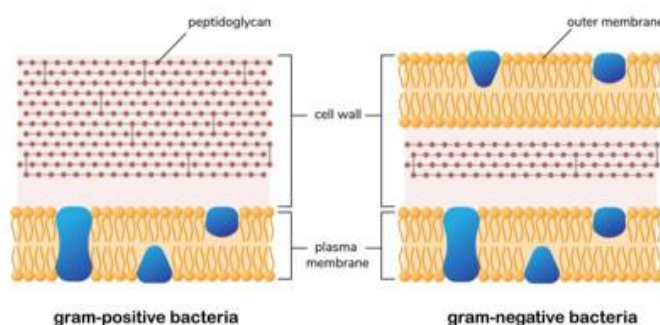


Figure 1. Gram-Positive versus Gram-Negative Bacteria Cell Wall.

The antimicrobial spectrum of *C. atropurpureus* spans Gram-positive bacteria, Gram-negative bacteria, fungi, and mycobacteria. Gram-positive bacteria have a simpler, single-layered peptidoglycan wall, while Gram-negative bacteria are protected by a complex outer membrane containing lipopolysaccharides, which acts as a formidable barrier against many antimicrobial agents. The density of the lipopolysaccharide in the outer bacterial cell wall layer (Figure 1) is much higher in the Gram-positive strain than Gram-negative strain. They are also found to suppress the growth of *M. tuberculosis* (Yanto et al. 2020; Marlina et al. 2021;

Pakadang et al. 2022; Rosamarlina et al. 2022). The compound abietane dipertenoids from *C. atropurpureus* possess anti-MRSA (methicillin-resistant *Staphylococcus aureus*) activity, showing inhibition zones observed at higher leaf extract concentrations fall within the "moderate to strong" activity range typically used in antimicrobial assays (i.e., >15 mm) (Jurkaninová et al. 2019; Mustarichie et al. 2022). Fungal pathogens such as *C. albicans* also show moderate susceptibility, broadening the spectrum of activity in opportunistic fungal infections.

When combined with conventional antibiotics such as ciprofloxacin and ampicillin, *C. atropurpureus* extracts showed synergistic effects. When its extracts are used with conventional antibiotics like ciprofloxacin and ampicillin, they can create a synergistic effect. This not only boosts the overall antimicrobial power but also allows for lower doses of the conventional drug. Findings also suggest that the leaf extract may serve as adjuvant therapy with anti-TB (Tuberculosis) drugs, such as rifampicin (Rosamarlina et al. 2021). This points to a potential role in combination therapy, reducing required doses of antibiotics and potentially delaying drug resistance.

Discussion

Mechanism of Action

Multiple studies confirm the presence of diverse bioactive compounds in *C. atropurpureus* leaves, including flavonoids (such as quercetin and luteolin), phenolic acids (such as rosmarinic acid), terpenoids (notably abietane-type diterpenoids), tannins, and saponins. These secondary metabolites contribute to the plant's antimicrobial activity through various mechanisms. Although direct mechanisms are not fully studied, suggested theories include disruption of microbial lipid membranes by terpenoids (Mahizan et al. 2019), enzyme inhibition by flavonoid-metal ion chelation (Kejík et al. 2021), reactive oxygen species (ROS) causing oxidative damage (Veiko et al. 2023), and bacteria DNA gyrase inhibition (Górniak et al. 2019). The multi-compound nature of the plant extract allows it to interfere with bacteria through several mechanisms simultaneously. By disrupting these various cellular functions, the active compounds extract can make the bacteria more vulnerable to the effects of a conventional antibiotic, effectively lowering the concentration of the drug needed to achieve a therapeutic effect.

C. atropurpureus may support antimicrobial activity by enhancing the host's immune response. During bacterial infections, inflammation and local tissue hypoxia can increase the expression of HIF-1 α a key regulator that boosts immune defense by promoting phagocytosis, nitric oxide synthase (NOS) activity, and the production of antimicrobial peptides

(Ramakrishnan et al. 2014; Yanto et al. 2020). These factors help directly eliminate pathogens. In addition, inflammation stimulates ICAM-1, which facilitates the movement of immune cells to the site of infection, further strengthening the body's defense against microbes (Bhalla et al. 2015).

Terpenoids (isoprenoids) are categorized as monoterpenes, diterpenoids, sesquiterpenes, and others. Terpenes have a lipophilicity tendency to penetrate and destroy bacteria cell wall, resulting in the difference in intra- and extracellular ATP concentration which disrupts the cell membrane, capsules, or biofilms, thus conducting the antibacterial activity (Nazzaro et al. 2013). In general, terpenoids exhibit substantial effects on Gram-positive and Gram-negative strains, with Gram-positive bacteria having greater antimicrobial vulnerability due to differences in membrane structure. They are also found to inhibit the quorum sensing communication system of bacteria coordination. The quorum sensing system is the main reason for the emergence of antibiotic resistance (Sharma et al. 2020). In addition, terpenoids also work as inhibitors of protein synthesis, and protein denaturing agents, which can achieve an antimicrobial effect.

In vitro study shows that antioxidants, such as flavonoids have been reported to enhance the number of CD4⁺ T-cells and increase the production of IFN- γ levels, a key cytokine involved in the activation of macrophages and the promotion of cellular immunity (Ullah et al. 2020; Pakadang et al. 2022). IFN- γ is critical in enhancing the antimicrobial functions of immune cells, including the upregulation of phagocytosis and the production of reactive oxygen and nitrogen species (Hosseinzade et al. 2019; Jomova et al. 2025). Flavonoids work by the methylation of the active hydroxyl groups generally eliminated or weakened the bacteria, by inhibiting their metabolism, disrupting the cell wall of bacteria, increasing its cell membrane permeability, and reducing the expression of virulence factors (Tran et al. 2012).

Tannins are a natural and diverse group of phenolic compounds that can bind to and clump together with proteins. (Xu et al. 2017; Farha et al. 2020) They are capable of binding with lipopolysaccharides and destabilize the integrity of the outer membrane and inhibit the biosynthesis of fatty acids.(Delehanty et al. 2007; Wu et al. 2010) Along with saponins, they may precipitate microbial proteins and interfere with membrane integrity or function.(Cui et al. 2018; Abd El-kader et al. 2020; Li and Monje-Galvan 2023) The results of an acute toxicity study indicate that the *C. atropurpureus* extract exhibits a favorable safety profile in mice, with no observed mortality up to a dose of 5000 mg/kg body weight. This suggests a positive indicator of the extract for further investigation into its safety for human use (Khattak et al. 2011).

Challenges and Future Prospects

Improvements in global health over recent decades are under threat because microorganisms that cause human diseases and medical conditions have become resistant to a wide range of antibiotics. Clinicians need to use last-resort medicines that are more costly, may have more side effects, and are often unavailable or unaffordable in low- and middle-income countries (Organization 2015). The development for new antibiotics has been alarmingly stagnant, with very few novel classes developed, which is a major concern because it allows for the emergence of drug-resistant “superbugs”. The misuse and overuse of existing antibiotics are primary drivers, both in healthcare settings and in agriculture. Particularly important gaps in knowledge that need to be filled are understanding how resistance develops in the environment (e.g. the specific pathways and genetic factors that allow bacteria to develop resistance), and exploring new sources of antimicrobial agents for novel drug discovery.

The main challenges of developing *C. atropurpureus* into a viable antimicrobial agent are the lack of active compounds identification and standardization, proving its safety and effectiveness through clinical trials, and ensuring a sustainable supply. *C. atropurpureus* extract contains a complex mix of secondary metabolites, but currently the specific compounds responsible for its antimicrobial activity are not fully identified or quantified. The lack of standardized extracts makes it challenging to ensure batch-to-batch consistency in terms of potency and therapeutic efficacy. Further research on isolating and identifying these specific compounds are also needed to understand their exact mechanisms of action.

While traditional use suggests the plant is safe (Quisumbing 1978; Roosita et al. 2008) research on toxicity studies are still lacking. To be approved as a potential medicine, testing of *C. atropurpureus* on animal cells to human cells are needed, especially with long term use. The effectiveness of the extracts compound in the human body is also unknown how well they are absorbed, metabolized, and delivered to the site of infection. Most research is at the basic science level. Previous in vitro studies may show strong antibacterial activity, but that does not guarantee the same result in a living organism (in vivo). Large scale controlled clinical studies are then needed to be compared to existing antibiotics. To transition from a folk remedy to a globally recognized medicine, future studies will need to move beyond in vitro lab experiments to large scale clinical human trials.

Despite the aforementioned challenges, future prospects on *C. atropurpureus* as a novel therapeutic agent are considerable through pharmacological approaches. The complex phytochemical profile of *C. atropurpureus* is its greatest strength, as it could provide a source for new drugs that use multi target mechanisms (unlike single compound antibiotics), which

would be harder for microorganisms to develop resistance to. This could then be used to combine purified plant compound with existing antibiotic. This approach provides a cost-effective and potentially less toxic alternative to developing entirely new synthetic drugs, which is a slow and expensive process.

4. KESIMPULAN

C. atropurpureus demonstrates promising antimicrobial activity against a broad spectrum of bacterial pathogens, largely attributed to its rich phytochemical content. Its extracts exhibit both standalone antimicrobial effects and synergistic potential when combined with standard antibiotics.

There is potential for *C. atropurpureus* to act as an adjuvant therapy alongside conventional antibiotics, however limitations are substantial. First, the limited number of in vivo studies and variability in methodologies across research (e.g. clinical trials) make it difficult to draw definitive conclusions regarding its clinical efficacy. Second, differences in methodologies, such as extraction method and microbial strains create variability that limits cross-study comparability. Third, data on pharmacokinetics, toxicity, and safe dosage are still lacking.

Further research is necessary to evaluate the safety, pharmacokinetics, and efficacy of standardized *C. atropurpureus* extracts in animal models and clinical trials. Future research should address these gaps by prioritizing standardization of extracts, such as identification of marker compounds for quality control. More in vivo studies are needed, followed by randomized clinical trials and toxicity studies. As resistance continues to rise, plant-based therapeutics should come to consideration.

DAFTAR REFERENSI

- Abd El-kader, A.M., Mahmoud, B.K., Hajjar, D., Mohamed, M.F., Hayallah, A.M. and Abdelmohsen, U.R. 2020. Antiproliferative activity of new pentacyclic triterpene and a saponin from *Gladiolus segetum* Ker-Gawl corms supported by molecular docking study. *RSC Advances* 10(38), pp. 22730-22741. <https://doi.org/10.1039/D0RA02775H>
- Arumugam, G., Swamy, M.K. and Sinniah, U.R. 2016. *Plectranthus amboinicus* (Lour.) Spreng: botanical, phytochemical, pharmacological and nutritional significance. *Molecules* 21, p. 369. <https://doi.org/10.3390/molecules21040369>
- Astuti, A.D., Yasir, B., Subehan and Alam, G. 2019. Comparison of two varieties of *Plectranthus scutellarioides* based on extraction method, phytochemical compound, and cytotoxicity. In: *Journal of Physics: Conference Series*. p.

72012.<https://doi.org/10.1088/1742-6596/1341/7/072012>

- Barbosa, M.O., Wilairatana, P., Leite, G.M., Delmondes, G.A., Silva, L.Y., Júnior, S.C. and others. 2023. *Plectranthus* species with anti-inflammatory and analgesic potential: a systematic review on ethnobotanical and pharmacological findings. *Molecules* 28, p. 5653.<https://doi.org/10.3390/molecules28155653>
- Bauer, N., Levanic, D.L., Mihaljevic, S. and Jelaska, S. 2002. Genetic transformation of *Coleus blumei* Benth. using *agrobacterium*. *Food Technology and Biotechnology* 40, pp. 163-169.
- Bhalla, K., Chugh, M., Mehrotra, S., Rathore, S. and Tousif, S. 2015. Host ICAMs play a role in cell invasion by *Mycobacterium tuberculosis* and *Plasmodium falciparum*. *Nature Communications* 6, p. 6049.<https://doi.org/10.1038/ncomms7049>
- Bismelah, N.A., Ahmad, R., Kassim, Z.H.M. and De Angelis, N. 2025. Antibacterial Activity of *Plectranthus scutellarioides* Leaf Against *Actinomyces viscosus* and Biocompatibility Testing on hFOB 1.19 Cell Line. *The Open Dentistry Journal* 19(1).<https://doi.org/10.2174/0118742106352620250416054519>
- Bismelah, N.A., Ahmad, R., Kassim, Z.M. and Ismail, N.H. 2019. *Coleus blumei* extract as a potential antibacterial oral rinse. In: *IOP Conference Series: Earth and Environmental Science*. p. 12020.<https://doi.org/10.1088/1755-1315/269/1/012015>
- Bismelah, N.A., Ahmad, R., Mohamed Kassim, Z.H., Ismail, N.H. and Rasol, N.E. 2022. The antibacterial effect of *Plectranthus scutellarioides* (L.) R.Br. leaves extract against bacteria associated with peri-implantitis. *Journal of Traditional and Complementary Medicine* 12(6), pp. 556-566.<https://doi.org/10.1016/j.jtcme.2022.07.002>
- Büttner, R. 2001. *Mansfeld's encyclopedia of agricultural and horticultural crops: (except ornamentals)*. Hanelt, P. ed. Berlin: Springer.
- Cui, C., Zong, J., Sun, Y., Zhang, L., Ho, C.T., Wan, X. and Hou, R. 2018. Triterpenoid saponins from the genus *Camellia*: structures, biological activities, and molecular simulation for structure-activity relationship. *Food & Function* 9(6), pp. 3069-3091.<https://doi.org/10.1039/C8FO00755A>
- DAR, K.B. 2016. Efficacy of Aqueous and Methanolic Extracts of *Rheum Spiciformis* against Pathogenic Bacterial and Fungal Strains. *JOURNAL OF CLINICAL AND DIAGNOSTIC RESEARCH*.<https://doi.org/10.7860/JCDR/2016/18036.8486>
- Delehanty, J.B., Johnson, B.J., Hickey, T.E., Pons, T. and Ligler, F.S. 2007. Binding and neutralization of lipopolysaccharides by plant proanthocyanidins. *Journal of Natural Products* 70(11), pp. 1718-1724.<https://doi.org/10.1021/np0703601>
- Devriese, E.G., Buffel, K. and Geuns, J.M. 1988. Stimulation of adventitious root formation on mung bean cuttings by coleon O. *Phytochemistry* 27(1), pp. 293-294.[https://doi.org/10.1016/0031-9422\(88\)80639-3](https://doi.org/10.1016/0031-9422(88)80639-3)
- Farha, A.K., Yang, Q.Q., Kim, G., Li, H.B., Zhu, F., Liu, H.Y. and others. 2020. Tannins as an alternative to antibiotics. *Food Bioscience* 38.<https://doi.org/10.1016/j.fbio.2020.100751>

- Górniak, I., Bartoszewski, R. and Króliczewski, J. 2019. Comprehensive review of antimicrobial activities of plant flavonoids. *Phytochemistry Reviews* 18, pp. 241-272. <https://doi.org/10.1007/s11101-018-9591-z>
- Hanum, F.S., Witaningrum, A.M. and Puspitasari, Y. [no date]. Effect of *Plectranthus scutellarioides* (L.) Leaf Extract as Natural Antibacterial Against *Staphylococcus aureus* and *Escherichia coli* Isolated From Dairy Cattle with Subclinical Mastitis. Available at: <https://e-journal.unair.ac.id/JBMV>.
- Hosseinzade, A., Sadeghi, O., Biregani, A.N., Soukhtehzari, S., Brandt, G.S. and Esmailzadeh, A. 2019. Immunomodulatory effects of flavonoids: possible induction of T CD4+ regulatory cells through suppression of mTOR pathway signaling activity. *Frontiers in Immunology* 10, p. 51. https://doi.org/10.1007/978-3-030-16073-9_22
- Ito, T., Rakainsa, S.K., Nisa, K. and Morita, H. 2018. Three new abietane-type diterpenoids from the leaves of Indonesian *Plectranthus scutellarioides*. *Fitoterapia* 127, pp. 146-150. <https://doi.org/10.1016/j.fitote.2018.02.013>
- Jomova, K., Alomar, S.Y., Valko, R., Liska, J., Nepovimova, E., Kuca, K. and others. 2025. Flavonoids and their role in oxidative stress, inflammation, and human diseases. *Chemico-Biological Interactions* 413, p. 111489. <https://doi.org/10.1016/j.cbi.2025.111489>
- Jurkaninová, S., Kubínová, R., Nejezchlebová, M., Gazdová, M., Hanáková, Z. and Dall'Acqua, S. 2019. Anti-MRSA activity of abietane diterpenes from *Coleus blumei* Benth. *Natural Product Research*. <https://doi.org/10.1080/14786419.2019.1686371>
- Karo, M., Hatta, M., Salma, W. and others. 2018. Effects of Miana (*Coleus scutellarioides* [L.] Benth) to expression of mRNA IL-37 in Balb/c mice infected *Candida albicans*. *Pharmacognosy Journal* 10, pp. 16-19. <https://doi.org/10.5530/pj.2018.1.3>
- Kaunang, V.M., Rares, R.W. and Sangari, F.J. 2016. Uji aktivitas antibakteri ekstrak daun mayana jantan (*Coleus atropurpureus* Benth) terhadap pertumbuhan bakteri *Streptococcus* sp. dan *Pseudomonas* sp. *e-Biomedik (eBm)* 4(1), pp. 489-494. <https://doi.org/10.35790/ebm.4.1.2016.10860>
- Kejík, Z. et al. 2021. Iron complexes of flavonoids-antioxidant capacity and beyond. *International Journal of Molecular Sciences* 22(2), p. 646. <https://doi.org/10.3390/ijms22020646>
- Khattak, M., Taher, M., Suzanah, A. and Ibrahim, A.B. 2011. Antibacterial and anti-fungal activity of coleus leaves consumed as breast milk stimulant. *International Journal of Nutrition and Food Sciences* 43(6), pp. 582-590. <https://doi.org/10.1108/NFS-11-2011-0131>
- Kowalczyk, T. et al. 2024. Biological Properties of Extracts Obtained from In Vitro Culture of *Plectranthus scutellarioides* in a Cell Model. *International Journal of Molecular Sciences* 25(2), p. 1043. <https://doi.org/10.3390/ijms25021043>
- Kubinová, R., Gazdová, M., Hanáková, Z., Jurkaninová, S., Dall'Acqua, S., Cvacka, J. and others. 2019. New diterpenoid glucoside and flavonoids from *Plectranthus scutellarioides* (L.) R. Br. *South African Journal of Botany* 120, pp. 286-

290. <https://doi.org/10.1016/j.sajb.2018.08.023>
- Lamprecht, W.J., Applegate, H. and Powell, R.D. 1975. Pigments of *Coleus blumei*. *Phytochemistry*.
- Levita, J., Sumiwi, S.A., Pratiwi, T.I., Ilham, E., Sidiq, S.P. and Moektiwardoyo, M. 2016. Pharmacological Activities of *Plectranthus scutellarioides* (L.) R.Br. leaves extract on cyclooxygenase and xanthine oxidase enzymes. *Journal of Medicinal Plants Research* 10, pp. 261-269.
- Li, J. and Monje-Galvan, V. 2023. In Vitro and In Silico Studies of Antimicrobial Saponins: A Review. *Processes* 11(10), p. 2856. <https://doi.org/10.3390/pr11102856>
- Lukhoba, C.W., Simmonds, M.S. and Paton, A.J. 2006. *Plectranthus*: A review of ethnobotanical uses. *Journal of Ethnopharmacology* 103, pp. 1-24. <https://doi.org/10.1016/j.jep.2005.09.011>
- Mahizan, N.A. et al. 2019. Terpene derivatives as a potential agent against antimicrobial resistance (AMR) pathogens. *Molecules* 24(14), p. 2631. <https://doi.org/10.3390/molecules24142631>
- Marlina, R., Hatta, M., Sridiana, E. and others. 2021. The effect of *Miana* (*Coleus scutellarioides* [L]) on vascular endothelial growth factor expression in Balb/c mice infected with *Mycobacterium tuberculosis*. *Biomedicine and Pharmacology Journal* 14, pp. 525-532. <https://doi.org/10.13005/bpj/2154>
- Moektiwardoyo, M., Levita, J., Sidiq, S.P., Ahmad, K., Mustarichie, R. and Subarnas, A. 2011. The determination of quercetin in *Plectranthus scutellarioides* (L.) R.Br. leaves extract and its in silico study on histamine H4 receptor. *Majalah Farmasi Indonesia* 22, pp. 191-196.
- Mu, Q., Zhang, H.J., Li, C.M. and Sun, H.D. 1996. A new diterpenoid from *Coleus scutellarioides*. *Phytochemistry*.
- Mustarichie, R., Iskandar, Y. and Saptarini, N.M. 2022. *Coleus atropurpureus* (L) Benth. Leaves as a New Promising Drug for Abscesses Caused by Methicillin-resistant *Staphylococcus aureus* and *Staphylococcus aureus*. *Pharmacognosy Journal* 14(2), pp. 439-443. <https://doi.org/10.5530/pj.2022.14.56>
- Namsa, N.D., Tag, H., Mandal, M., Kalita, P. and Das, A.K. 2009. An ethnobotanical study of traditional anti-inflammatory plants used by the Lohit community of Arunachal Pradesh, India. *Journal of Ethnopharmacology* 125(2), pp. 234-245. <https://doi.org/10.1016/j.jep.2009.07.004>
- Nazzaro, F., Fratianni, F., De Martino, L., Coppola, R. and De Feo, V. 2013. Effect of Essential Oils on Pathogenic Bacteria. *Pharmaceuticals* 6(12), pp. 1451-1474. <https://doi.org/10.3390/ph6121451>
- Nguyen, P. and Cin, V.D. 2009. The role of light on foliage colour development in *coleus* (*Solenostemon scutellarioides* (L.) Codd). *Plant Physiology and Biochemistry* 47(10), pp. 934-945. <https://doi.org/10.1016/j.plaphy.2009.06.006>

- Organization, W.H. 1978. The promotion and development of traditional medicine: report of a WHO meeting held in Geneva from 28 November to 2 December 1977. Geneva: World Health Organization.
- Organization, W.H. 2015. Global action plan on antimicrobial resistance. Geneva: World Health Organization.
- Pakadang, S.R., Ratnah, S., Salasa, A.M. and others. 2022. Toll like receptor 4 expression profile in mice infected Mycobacterium tuberculosis given with miana leaves extract (*Coleus scutellarioides* [L.] Benth). Biomedicine and Pharmacology Journal. <https://doi.org/10.5530/pj.2022.14.63>
- Paton, A.J., Springate, D., Suddee, S., Otieno, D., Grayer, R.J., Harley, M.M. and others. 2004. Phylogeny and evolution of basils and allies (Ocimeae, Labiatae) based on three plastid DNA regions. Molecular Phylogenetics and Evolution 31(1), pp. 277-299. <https://doi.org/10.1016/j.ympev.2003.08.002>
- Plants of the World Online. 2023. *Coleus scutellarioides* (L.) Benth.
- Quattrocchi, U. 2012. CRC world dictionary of medicinal and poisonous plants. Cleveland: CRC Press.
- Quisumbing, E. 1978. Medicinal plants of the Philippines. Manila: Bureau of Printing.
- Ragasa, C.Y., Templora, V.F. and Rideout, J.A. 2001. Diastereomeric diterpenes from *Coleus blumei*. Chemical and Pharmaceutical Bulletin 49(7), pp. 927-929. <https://doi.org/10.1248/cpb.49.927>
- Ramakrishnan, S., Anand, V. and S, R. 2014. Vascular endothelial growth factor signaling in hypoxia and inflammation. Journal of Neuroimmune Pharmacology 9(2), pp. 42-60. <https://doi.org/10.1007/s11481-014-9531-7>
- Ridwan, Y. and Ayunita, Y.Q. 2007. Phytochemical and anthelmintic activity against chicken tapeworm of painted nettle (*Coleus blumei* Benth.) varieties in vitro. J Protein 14, pp. 17-20.
- Roosita, K., Kusharto, C.M., Sekiyama, M., Fachrurrozi, Y. and Ohtsuka, R. 2008. Medicinal plants used by the villagers of a Sundanese community in West Java, Indonesia. Journal of Ethnopharmacology 115(1), pp. 72-81. <https://doi.org/10.1016/j.jep.2007.09.010>
- Rosamarlina, R., Hatta, M., Djaharuddin, I. and others. 2022. The changes of HIF-1 α and ICAM-1 expression after Miana (*Coleus scutellarioides* [L.]) treatment in Balb/C mice with Mycobacterium tuberculosis infection. Biomedicine and Pharmacology Journal 15, pp. 73-81. <https://doi.org/10.13005/bpj/2344>
- Rosamarlina, R., Hatta, M., Sridiana, E., Djaharuddin, I., Patellongi, I. and Murtian, E. 2021. The effect of Miana (*Coleus scutellarioides* [L.]) on Vascular Endothelial Growth Factor expression in Balb/c mice infected with Mycobacterium tuberculosis. Biomedicine and Pharmacology Journal 14, pp. 525-532. <https://doi.org/10.13005/bpj/2154>
- Sharma, A., Biharee, A., Kumar, A. and Jaitak, V. 2020. Antimicrobial Terpenoids as a Potential Substitute in Overcoming Antimicrobial Resistance. Current Drug Targets

- 21(14), pp. 1476-1494.<https://doi.org/10.2174/1389450121666200520103427>
- Thomas, S. 2006. Taiwanese native medicinal plants: phytopharmacology and therapeutic values. New York: CRC Press.
- Tran, T.-D., Do, T.-H., Tran, N.-C., Ngo, T.-D., Huynh, T.-N.-P., Tran, C.-D. and Thai, K.-M. 2012. Synthesis and anti Methicillin resistant *Staphylococcus aureus* activity of substituted chalcones alone and in combination with non-beta-lactam antibiotics. *Bioorganic & Medicinal Chemistry Letters* 22(14), pp. 4555-4560.<https://doi.org/10.1016/j.bmcl.2012.05.112>
- Ullah, A. et al. 2020. Important Flavonoids and Their Role as a Therapeutic Agent. *Molecules* 25(22), p. 5243.<https://doi.org/10.3390/molecules25225243>
- Vaou, N., Stavropoulou, E., Voidarou, C., Tsigalou, C. and Bezirtzoglou, E. 2021. Towards advances in medicinal plant antimicrobial activity: a review study on challenges and future perspectives. *Microorganisms* 9(10), p. 2041.<https://doi.org/10.3390/microorganisms9102041>
- Veiko, A.G., Olchowik-Grabarek, E., Sekowski, S., Roszkowska, A., Lapshina, E.A., Dobrzynska, I. and others. 2023. Antimicrobial activity of quercetin, naringenin and catechin: flavonoids inhibit *Staphylococcus aureus*-induced hemolysis and modify membranes of bacteria and erythrocytes. *Molecules* 28(3), p. 1252.<https://doi.org/10.3390/molecules28031252>
- Wiar, C. 2006. Medicinal plants of the Asia-Pacific: drugs for the future? Singapore: World Scientific.<https://doi.org/10.1142/9789812707260>
- Wu, D., Wu, X.D., You, X.F., Ma, X.F. and Tian, W.X. 2010. Inhibitory effects on bacterial growth and β -ketoacyl-ACP reductase by different species of maple leaf extracts and tannic acid. *Phytotherapy Research* 24(S1), pp. S35-S41.<https://doi.org/10.1002/ptr.2873>
- Xu, Y., Shi, C., Wu, Q., Zheng, Z., Liu, P., Li, G. and others. 2017. Antimicrobial activity of punicalagin against *Staphylococcus aureus* and its effect on biofilm formation. *Foodborne Pathogens and Disease* 14, pp. 282-287.<https://doi.org/10.1089/fpd.2016.2226>
- Yanto, T.A., Hatta, M., Bukhari, A. and Natzir, R. 2020. Molecular and immunological mechanisms of Miana leaf (*Coleus scutellarioides* [L.] Benth) in infectious diseases. *Biomedical and Pharmacology Journal* 13(4), pp. 1607-1618.<https://doi.org/10.13005/bpj/2036>
- Zakaria, Z., Aziz, R., Lachimanan, Y.L., Sreenivasan, S. and Rathinam, X. 2008. Antioxidant activity of *Coleus blumei*, *Orthosiphon stamineus*, *Ocimum basilicum* and *Mentha arvensis* from Lamiaceae family. *IJNES* 2, pp. 93-95.